

AMP-activated Protein Kinase $\alpha 2$ Subunit Is Required for the Preservation of Hepatic Insulin Sensitivity by n-3 Polyunsaturated Fatty Acids

Tomas Jelenik,¹ Martin Rossmeisl,¹ Ondrej Kuda,¹ Zuzana Macek Jilkova,¹ Dasa Medrikova,¹ Vladimir Kus,¹ Michal Hensler,¹ Petra Janovska,¹ Ivan Miksik,² Marcin Baranowski,³ Jan Gorski,³ Sophie Hébrard,^{4,5} Thomas E. Jensen,⁶ Pavel Flachs,¹ Simon Hawley,⁷ Benoit Viollet,^{4,5} and Jan Kopecky¹

OBJECTIVE—The induction of obesity, dyslipidemia, and insulin resistance by high-fat diet in rodents can be prevented by n-3 long-chain polyunsaturated fatty acids (LC-PUFAs). We tested a hypothesis whether AMP-activated protein kinase (AMPK) has a role in the beneficial effects of n-3 LC-PUFAs.

RESEARCH DESIGN AND METHODS—Mice with a whole-body deletion of the $\alpha 2$ catalytic subunit of AMPK (AMPK $\alpha 2^{-/-}$) and their wild-type littermates were fed on either a low-fat chow, or a corn oil-based high-fat diet (cHF), or a cHF diet with 15% lipids replaced by n-3 LC-PUFA concentrate (cHF+F).

RESULTS—Feeding a cHF diet induced obesity, dyslipidemia, hepatic steatosis, and whole-body insulin resistance in mice of both genotypes. Although cHF+F feeding increased hepatic AMPK $\alpha 2$ activity, the body weight gain, dyslipidemia, and the accumulation of hepatic triglycerides were prevented by the cHF+F diet to a similar degree in both AMPK $\alpha 2^{-/-}$ and wild-type mice in ad libitum-fed state. However, preservation of hepatic insulin sensitivity by n-3 LC-PUFAs required functional AMPK $\alpha 2$ and correlated with the induction of adiponectin and reduction in liver diacylglycerol content. Under hyperinsulinemic-euglycemic conditions, AMPK $\alpha 2$ was essential for preserving low levels of both hepatic and plasma triglycerides, as well as plasma free fatty acids, in response to the n-3 LC-PUFA treatment.

CONCLUSIONS—Our results show that n-3 LC-PUFAs prevent hepatic insulin resistance in an AMPK $\alpha 2$ -dependent manner and support the role of adiponectin and hepatic diacylglycerols in the regulation of insulin sensitivity. AMPK $\alpha 2$ is also essential for hypolipidemic and antisteatotic effects of n-3 LC-PUFA under insulin-stimulated conditions. *Diabetes* 59:2737–2746, 2010

From the ¹Department of Adipose Tissue Biology and the ²Department of Analysis of Biologically Important Compounds, Institute of Physiology of the Academy of Sciences of the Czech Republic, v.v.i., Prague, Czech Republic; the ³Department of Physiology, Medical University of Bialystok, Poland; the ⁴Institut Cochin, Université Paris Descartes, CNRS (UMR 8104), Paris, France; ⁵INSERM, U1016, Paris, France; the ⁶Molecular Physiology Group, Copenhagen Muscle Research Centre, Department of Exercise and Sport Sciences, Section of Human Physiology, University of Copenhagen, Copenhagen, Denmark; and the ⁷Division of Molecular Physiology, College of Life Sciences, University of Dundee, Scotland, U.K.

Corresponding author: Jan Kopecky, kopecky@biomed.cas.cz.

Received 20 November 2009 and accepted 26 July 2010. Published ahead of print at <http://diabetes.diabetesjournals.org> on 6 August 2010. DOI: 10.2337/db09-1716.

T.J. and M.R. contributed equally to this study.

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Naturally occurring n-3 long-chain polyunsaturated fatty acids (LC-PUFAs)—namely, eicosapentaenoic acid (20:5n-3) and docosahexaenoic acid (22:6n-3)—which are abundant in sea fish, act as hypolipidemics, reduce cardiac events, and decrease the progression of atherosclerosis [reviewed in refs (1,2)]. Studies of obese humans have also demonstrated a reduction in adiposity after n-3 LC-PUFA supplementation (3,4). In rodents fed a high-fat diet, n-3 LC-PUFAs efficiently prevented the development of obesity, hepatic steatosis, and dyslipidemia (5–8), as well as impaired glucose tolerance (8–10). However, in diabetic patients, n-3 LC-PUFAs appear to have little effect on glycemic control (3,11,12).

The hypolipidemic and antiobesity effects of n-3 LC-PUFAs depend on both the suppression of lipogenesis and the increase in fatty acid oxidation in several tissues, including the liver (13,14), adipose tissue (6), and intestine (15). This metabolic switch may reduce the accumulation of toxic fatty acid derivatives, while protecting insulin signaling in the liver and muscle (9,10,16). Our previous work has documented that the preservation of whole-body insulin sensitivity by n-3 LC-PUFAs in mice fed a high-fat diet mainly reflects improved hepatic insulin sensitivity (8). The effects of n-3 LC-PUFAs and their active metabolites (17,18) are mediated by peroxisome proliferator-activated receptors (PPAR), with PPAR- α and PPAR- δ (β) being the main targets (14,16), although PPAR- γ , liver X receptor- α , hepatic nuclear factor-4, sterol regulatory element binding protein-1c (SREBP-1c) and carbohydrate-responsive element-binding protein are also involved (16,19–21).

It has been demonstrated that n-3 LC-PUFAs enhanced AMP-activated protein kinase (AMPK) activity in the liver (22), intestine (23), and adipose tissue (18,24). AMPK is a heterotrimeric protein consisting of a catalytic α -subunit and regulatory β - and γ -subunits, with multiple isoforms identified for each subunit [$\alpha 1$, $\alpha 2$, $\beta 1$, $\beta 2$, $\gamma 1$, $\gamma 2$, and $\gamma 3$; reviewed in ref (25)]. Experiments using whole-body AMPK $\alpha 2$ null [AMPK $\alpha 2^{-/-}$; ref (26)] mice showed the importance of the AMPK $\alpha 2$ subunit for whole-body insulin action, while liver-specific AMPK $\alpha 2$ knockout mice (27) as well as adenovirus-mediated activation of AMPK $\alpha 2$ in the liver (28) implicated the hepatic AMPK $\alpha 2$ isoform in the suppression of hepatic glucose production and maintenance of fasting blood glucose levels. Furthermore, AMPK controls metabolic fluxes in response to changing cellular

energy levels, namely, the partitioning between lipid oxidation and lipogenesis (29,30).

We hypothesized that the effects of n-3 LC-PUFA on insulin sensitivity and lipid metabolism in mice fed an obesogenic high-fat diet require a functional AMPK α 2 isoform. To test this hypothesis *in vivo*, AMPK α 2^{-/-} and wild-type mice were fed either a low-fat chow diet (Chow), a corn oil-based high-fat (cHF) diet, or cHF diet in which 15% of the lipids were replaced by n-3 LC-PUFA concentrate (cHF+F). Our results demonstrate an AMPK α 2-dependent action of n-3 LC-PUFAs, in 1) the preservation of hepatic and muscle insulin sensitivity; 2) the changes in hepatic diacylglycerol content and composition; and 3) the antisteatotic effect in the liver and hypolipidemic effect under insulin-stimulated conditions, such as during hyperinsulinemic-euglycemic clamp, but not when the organism depends on lipids as substrates.

RESEARCH DESIGN AND METHODS

Four-month-old whole-body AMPK α 2^{-/-} mice (29) backcrossed to C57BL/6J mice for nine generations, and wild-type littermate controls were fed on either Chow, cHF, or cHF+F diet for nine weeks. Body weight and food consumption were recorded, and EDTA-plasma and tissues were collected for various analyses as described in the online appendix, available at <http://diabetes.diabetesjournals.org/cgi/content/full/db09-1716/DC1>. Male mice were used for all the experiments, except for the measurements of hepatic AMPK activity, which were performed on female mice. The experiments were conducted under the guidelines for the use and care of laboratory animals of the Institute of Physiology and followed the "Principles of laboratory animal care" (National Institutes of Health publication no. 85-23, revised 1985).

Blood and plasma parameters. Blood glucose was measured using calibrated glucometers (LifeScan, Milpitas, CA). Nonesterified fatty acids (NEFAs), triglycerides, and total cholesterol were determined in plasma using the following enzymatic photometric tests: NEFA-C (Wako Chemicals, Neuss, Germany), triacylglycerols liquid, and cholesterol liquid (Pliva-Lachema Diagnostika, Brno, Czech Republic), respectively. Plasma insulin was measured using the Sensitive Rat Insulin RIA Kit (LINCO Research, St. Charles, MO). Total adiponectin levels and adiponectin multimeric complexes were determined using Western blotting (31).

Lipid content and gene expression in the liver. The tissue content of triglycerides was estimated in ethanolic KOH tissue lysates as described before (8). The content and fatty acid composition of the phospholipid, diacylglycerol, triglyceride, and ceramide fractions were assessed in tissue lipid extracts; gene expression was evaluated using real-time RT-PCR (see online appendix).

Activity of α 1 and α 2 AMPK isoforms. Livers were collected by freeze-clamping, AMPK was immunoprecipitated from tissue extracts, and the activity was assayed using a peptide substrate (32); see the online appendix.

Hyperinsulinemic-euglycemic clamp. Five days before the experiment, an indwelling catheter was placed into the left femoral vein under anesthesia (33). Mice were allowed to recover for 5–7 days, followed by a 6-h fast (8:00 A.M.–2:00 P.M.) prior to the experiment. The whole-body glucose turnover was determined under basal (nonstimulated) and insulin-stimulated conditions (hyperinsulinemic-euglycemic clamp), using separate groups of mice. Insulin (Actrapid, Novo Nordisk Pharma, Denmark) was infused at a constant rate of 4.8 mU/kg-min for 3 h, while D-[3-³H]glucose (Perkin Elmer, Boston, MA) was infused at a rate of 15.9 kBq/min. Throughout the infusion, glucose concentration and D-[3-³H]glucose specific activity (during the last hour of infusion) were determined in tail blood. Euglycemia (~5.55 mmol/l) was maintained by periodically adjusting a variable infusion of 33% glucose (33). At the end of a 3-h infusion period, mice were first anesthetized by diethylether, exsanguinated through the cervical incision, and then killed by cervical dislocation, and tissues (liver and quadriceps muscle) and EDTA-plasma were collected for biochemical analyses (see supplementary Table 1 and Research Design and Methods of the online appendix for details on basic clamp parameters and methodology).

Primary cultures of hepatocytes. Hepatocytes were isolated from livers of fed mice by a modification of the collagenase method (34) and seeded at a density of 0.5×10^6 cells per each 35-mm Petri dish. Rates of basal and insulin-stimulated *de novo* lipogenesis and AICAR-stimulated fatty acid oxidation were measured using [1-¹⁴C] acetate and [1-¹⁴C] palmitate, respectively (see online appendix).

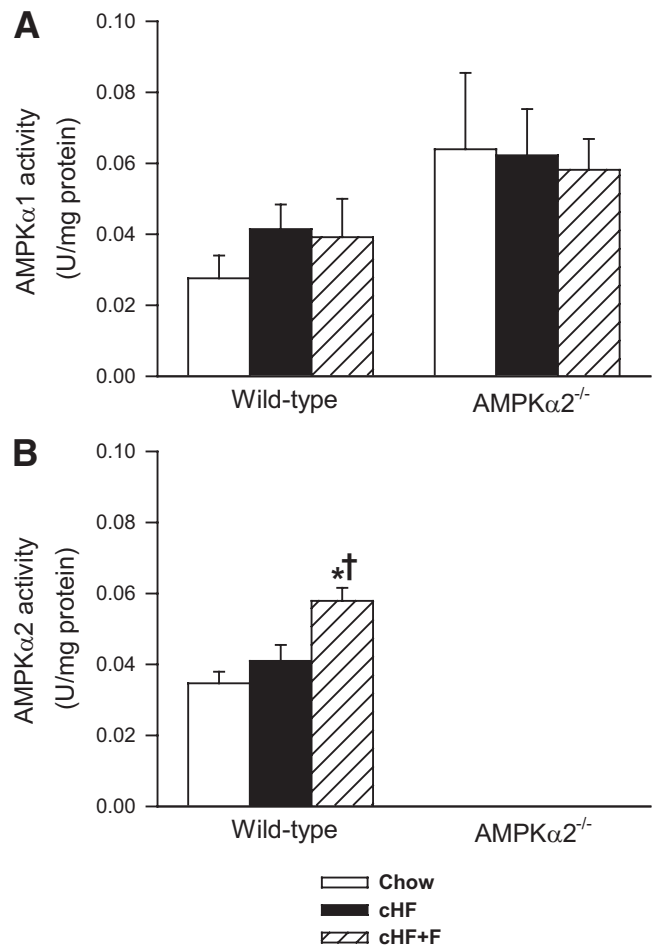


FIG. 1. Liver AMPK α 1 (A) and AMPK α 2 (B) activity in wild-type and AMPK α 2^{-/-} mice fed either a Chow diet, cHF, or cHF+F for 9 weeks. The data are the means \pm SE ($n = 5-8$). In the AMPK α 2^{-/-} mice, AMPK α 2 activity was below the detection limit. * $P < 0.05$ versus genotype Chow; † $P < 0.05$ versus genotype cHF.

Statistics. All values are presented as means \pm SE. Data were analyzed by two-way ANOVA. Comparisons were judged to be significant at $P \leq 0.05$.

RESULTS

Enhancement of hepatic AMPK α 2 activity by n-3 LC-PUFAs. Specific activities of AMPK α 1 and AMPK α 2 were evaluated in the liver of ad libitum-fed mice after nine weeks of the differential dietary treatment (Fig. 1). No significant effect of either diet (Chow, cHF, and cHF+F) or genotype (wild-type versus AMPK α 2^{-/-}) on AMPK α 1-specific activity was observed, although the AMPK α 1 activity tended to be higher in the AMPK α 2^{-/-} mice (Fig. 1A). In contrast, AMPK α 2 activity was stimulated by n-3 LC-PUFAs (cHF+F diet; Fig. 1B). AMPK α 2 activity was not detected in the AMPK α 2^{-/-} mice (Fig. 1B). No changes were detected in the activity of AMPK α 1 and AMPK α 2 in the quadriceps muscle in response to n-3 LC-PUFAs (not shown).

AMPK α 2 is not required for antiobesity and hypolipidemic effects of n-3 LC-PUFAs in ad libitum-fed mice. At four months of age, at the beginning of dietary treatments, wild-type and AMPK α 2^{-/-} mice fed the Chow diet exhibited similar body weights (Table 1). In mice of both genotypes, cHF-feeding for nine weeks resulted in greater body weight gain compared with the Chow-fed mice. However, this effect was less pronounced in AMPK α 2^{-/-}

TABLE 1
Metabolic and plasma parameters in wild-type and AMPK α 2^{-/-} mice

Metabolic parameters	Wild-type			AMPK α 2 ^{-/-}		
	Chow	cHF	cHF+F	Chow	cHF	cHF+F
Food consumption (kJ/g · day)	2.0 ± 0.1	1.8 ± 0.1	1.9 ± 0.0	2.0 ± 0.1	1.8 ± 0.1	2.0 ± 0.1
Initial body weight (g)	27.8 ± 0.5	27.9 ± 0.5	27.7 ± 0.5	27.1 ± 0.3	27.7 ± 0.4	27.3 ± 0.4
Body weight gain (g)	1.4 ± 0.2	7.0 ± 1.1*	2.8 ± 0.7†	1.8 ± 0.3	4.6 ± 0.7*‡	1.2 ± 0.4†
Adiposity						
Epididymal AT (g)	0.43 ± 0.02	1.52 ± 0.19*	1.12 ± 0.14*†	0.37 ± 0.03	1.19 ± 0.16*‡	0.64 ± 0.07*†‡
Subcutaneous AT (g)	0.20 ± 0.01	0.54 ± 0.05*	0.42 ± 0.04*†	0.17 ± 0.01	0.34 ± 0.03*‡	0.23 ± 0.01†‡
Epididymal adipocytes size (μm ²)	ND	15,971 ± 1,784	10,232 ± 185†	ND	13,298 ± 1,632	8 593 ± 896†
Subcutaneous adipocytes size (μm ²)	2,916 ± 610	6,775 ± 1,718*	7,311 ± 1,308*	3,395 ± 139	6,625 ± 926*	5,432 ± 648*
Plasma metabolites						
Triglycerides (mmol/l)	1.17 ± 0.08	1.23 ± 0.11	0.62 ± 0.07*†	1.04 ± 0.06	1.22 ± 0.08	0.73 ± 0.06*†
NEFAs (mmol/l)	0.90 ± 0.05	0.94 ± 0.05	0.59 ± 0.04*†	0.88 ± 0.04	0.99 ± 0.06	0.68 ± 0.04*†
Cholesterol (mmol/l)	2.25 ± 0.08	4.12 ± 0.25*	3.10 ± 0.20*†	2.10 ± 0.06	3.94 ± 0.18*	2.75 ± 0.14*†
Glucose (mmol/l)	9.9 ± 0.4	10.7 ± 0.5	10.4 ± 0.3	9.7 ± 0.4	10.0 ± 0.4	9.4 ± 0.4
Plasma hormones						
Insulin fed (ng/ml)	0.66 ± 0.11	1.73 ± 0.29*	1.47 ± 0.28*	0.60 ± 0.07	1.34 ± 0.20*	0.95 ± 0.12*
Insulin fasted (ng/ml)	0.13 ± 0.01	0.39 ± 0.06*	0.19 ± 0.05†	0.16 ± 0.02	0.22 ± 0.03‡	0.15 ± 0.01
Total adiponectin (A.U.)	1.15 ± 0.09	0.97 ± 0.10	1.33 ± 0.09†	0.82 ± 0.09‡	0.80 ± 0.07	0.97 ± 0.08‡
HMW: total adiponectin	0.38 ± 0.02	0.36 ± 0.02	0.44 ± 0.02†	0.35 ± 0.02	0.34 ± 0.02	0.39 ± 0.02

Data are means ± SE of 27–30 mice for metabolic parameters and 13–15 mice for other measures. AMPK α 2^{-/-} and wild-type mice were fed either a Chow diet, cHF, or cHF+F for nine weeks. Food consumption was measured weekly for nine weeks. Body weight gain (see supplementary Fig. 1) and plasma parameters were assessed in ad libitum-fed mice after nine weeks. Plasma insulin levels were also assessed in fasted mice after eight weeks. AT, adipose tissue; A.U., arbitrary units; HMW, total adiponectin, ratio of high molecular weight to total adiponectin (for levels of all molecular weight forms of adiponectin; ND, no data; see supplementary Fig. 2); **P* < 0.05 vs. genotype Chow; †*P* < 0.05 vs. genotype cHF; ‡*P* < 0.05 vs. wild-type on respective diet.

mice (Table 1). In both wild-type and AMPK α 2^{-/-} mice, the cHF+F diet induced smaller body weight gain than the cHF diet (Table 1 and supplementary Figure 1). None of the differences in body weight gain could be explained by caloric intake, which was similar in all experimental groups (Table 1). The weight of fat depots increased in response to cHF feeding, while the cHF+F diet partially prevented this increase (Table 1). Triglycerides and NEFA levels in plasma of ad libitum-fed mice were similar in the Chow- and cHF-fed mice, while cholesterol levels were markedly and significantly elevated by the cHF diet. n-3 LC-PUFAs lowered plasma lipid levels independently of AMPK α 2. Triglycerides and NEFA levels were strongly reduced, even below the levels observed in the Chow-fed mice (Table 1).

AMPK α 2 is essential for the preservation of insulin sensitivity in response to n-3 LC-PUFAs. After nine weeks of dietary treatment, no change was observed in blood glucose, but elevations were observed in plasma insulin levels in response to the cHF diet in ad libitum-fed mice of both genotypes. However, the increase in plasma insulin levels was less pronounced in AMPK α 2^{-/-} mice, closely reflecting the genotype-dependent differences in body weight gain (Table 1). A similar pattern of changes in insulin levels was also observed in fasted mice, in which n-3 LC-PUFAs significantly reduced insulin levels only in wild-type animals (Table 1). As expected, plasma levels of total as well as high molecular weight form of adiponectin, an adipokine associated with increased insulin sensitivity (35), were increased ~1.4- and ~1.2-fold, respectively, in wild-type mice in response to n-3 LC-PUFA supplementation (Table 1 and supplementary Fig. 2, available in an online appendix); however, no significant increase of plasma adiponectin by n-3 LC-PUFAs was observed in AMPK α 2^{-/-} mice.

In further experiments, hyperinsulinemic-euglycemic

clamps were performed to evaluate whole-body insulin sensitivity. Under basal conditions, glucose turnover rate (GTO; i.e., glucose uptake in peripheral tissues) was similar in all groups of mice (supplementary Table 1). Under insulin-stimulated conditions (Fig. 2A–F), the amount of exogenous glucose required to maintain euglycemia during the clamp, i.e., the glucose infusion rate (GIR), was ~1.3-fold lower in AMPK α 2^{-/-} than in wild-type mice fed the Chow diet (Fig. 2A). On the other hand, GIR was decreased by the cHF diet to a similar level in mice of both genotypes, manifesting diet-induced insulin resistance. This was attributed to a decreased GTO and, in particular, to an impaired suppression of hepatic glucose production (HGP) by insulin, with HGP being ~8.5-fold higher in the cHF-fed compared with the Chow-fed wild-type mice (Fig. 2C). In wild-type mice, cHF+F diet feeding increased GIR and GTO (~1.9- and ~1.2-fold increase, respectively) as compared with cHF-fed mice, while HGP was lowered to a similar level as in the Chow-fed mice. These results document the protective effects of n-3 LC-PUFAs from high-fat diet-induced insulin resistance in wild-type mice, namely, at the level of HGP. In contrast, none of these beneficial effects of n-3 LC-PUFAs was observed in AMPK α 2^{-/-} mice, in which neither the GIR (Fig. 2A) nor the GTO (Fig. 2B) differed between the cHF+F-fed and the cHF-fed mice, whereas the rate of HGP was even higher in the cHF+F-fed than in the cHF-fed AMPK α 2^{-/-} mice (Fig. 2C). Although whole-body glycolysis was similar in all the groups (Fig. 2D), the rate of whole-body glycogen synthesis, which reflects insulin sensitivity of muscle glucose metabolism, was dependent on both diet and genotype (Fig. 2E). In the Chow-fed mice, the rate of whole-body glycogen synthesis tended to be higher in wild-type mice than in AMPK α 2^{-/-} mice. Only in the former mice was it significantly affected by dietary treatment. Thus, in wild-type mice, glycogen synthesis was

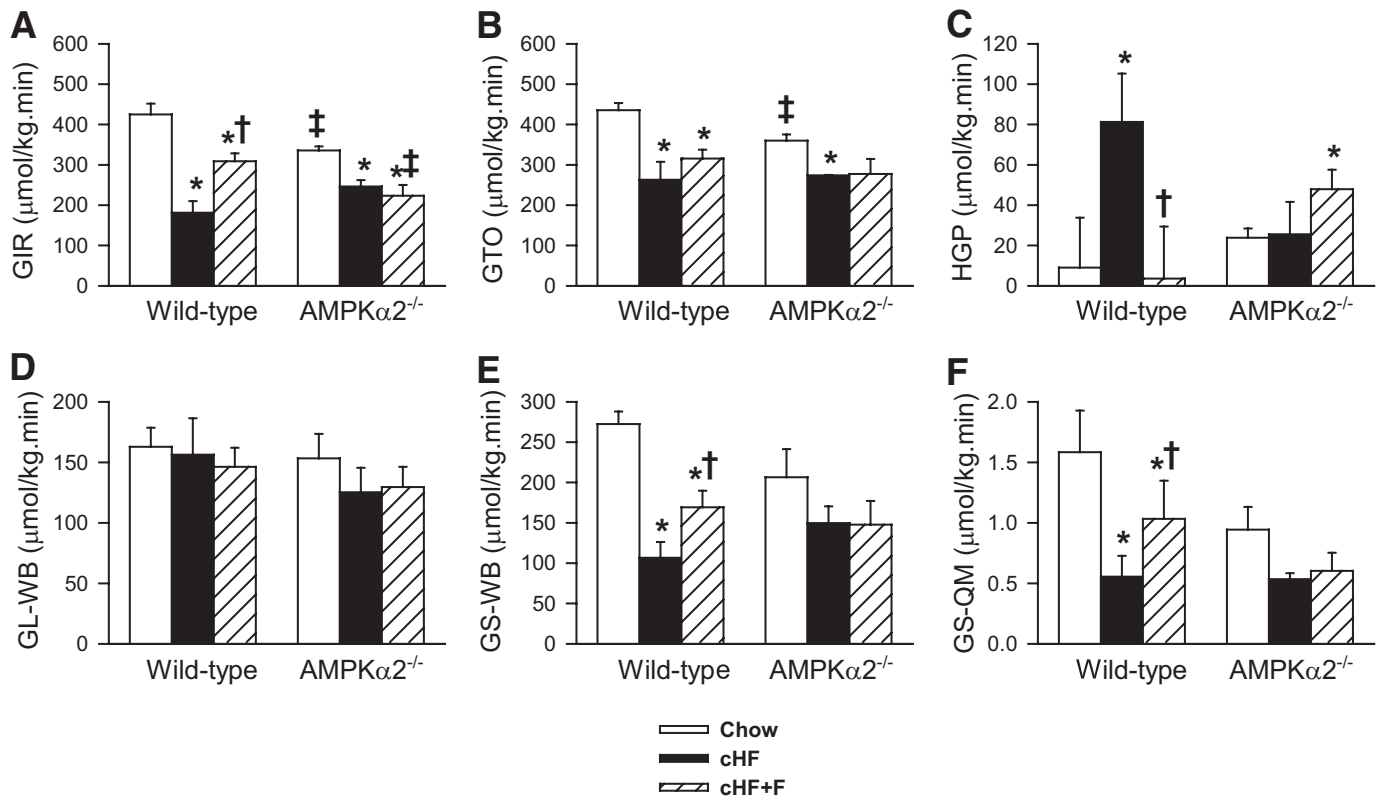


FIG. 2. Insulin sensitivity assessed by hyperinsulinemic-euglycemic clamp. GIR (A), GTO (B), HGP (C), whole-body glycolysis (GL-WB; D), whole-body glycogen synthesis (GS-WB; E); and glycogen synthesis in quadriceps muscle (GS-QM; F) were measured in wild-type and AMPK α 2^{-/-} mice fed either a Chow diet, cHF, or cHF+F for 9 weeks. The data are the means \pm SE ($n = 5-8$). * $P < 0.05$ versus genotype Chow; † $P < 0.05$ versus genotype cHF; ‡ $P < 0.05$ versus wild-type on respective diet.

decreased ~ 2.4 -fold in response to the cHF diet, while n-3 LC-PUFAs provided a partial protection from this decrease (Fig. 2E). A similar pattern of changes in the glycogen synthesis rate in response to n-3 LC-PUFAs was observed when measured directly in the skeletal muscle (Fig. 2F). Thus, in accordance with the previous study (26), the results of clamp studies suggested impairment of insulin sensitivity in response to whole-body ablation of AMPK α 2 in Chow-fed mice. However, AMPK α 2^{-/-} mice seemed to be partially protected against cHF-induced insulin resistance, while AMPK α 2 was required for preservation of insulin sensitivity in the skeletal muscle and especially in the liver in response to n-3 LC-PUFA feeding.

Unmasking the role of AMPK α 2 in the lipid-lowering effect of n-3 LC-PUFAs under hyperinsulinemic-euglycemic conditions. In addition to the ad libitum-fed mice (Table 1), plasma lipid levels were also measured in fasted mice, as well as in mice subjected to hyperinsulinemic-euglycemic clamp (supplementary Table 2). In contrast to the ad libitum-fed state, cHF+F diet did not affect either triglyceride or NEFA levels under fasting conditions. Under the hyperinsulinemic-euglycemic conditions, both triglyceride and NEFA levels were lower in the cHF+F-fed than in the cHF-fed wild-type mice (~ 1.6 -fold and ~ 1.4 -fold difference, respectively), but no such difference between the diets was observed in AMPK α 2^{-/-} mice. Cholesterol levels were consistently decreased by n-3 LC-PUFAs independently of both the metabolic state and genotype (supplementary Table 2).

In ad libitum-fed mice of both genotypes, the hepatic triglyceride content was increased \sim twofold by cHF compared with the Chow diet, while triglyceride accumulation

was increased only ~ 1.3 -fold by cHF+F diet in both genotypes, documenting a protection against hepatic triglyceride accumulation by n-3 LC-PUFAs (Fig. 3A). Under hyperinsulinemic-euglycemic conditions, n-3 LC-PUFAs also protected livers of wild-type mice against the cHF-induced accumulation of triglycerides. However, this effect was absent in AMPK α 2^{-/-} mice (Fig. 3B). Moreover, a strong correlation was found between plasma NEFA levels and hepatic triglyceride content assessed under the clamp conditions in the cHF+F-fed AMPK α 2^{-/-} mice ($R^2 = 0.43$, $P < 0.05$) but not in wild-type mice ($R^2 = 0.08$, $P = 0.40$).

Dietary n-3 LC-PUFAs increase 5-aminoimidazole-4-carboxamide-1- β -D-ribofuranoside-stimulated fatty acid oxidation and insulin-stimulated lipogenesis in cultured hepatocytes from wild-type but not from AMPK α 2^{-/-} mice. We sought to determine whether the differential effect of n-3 LC-PUFAs on accumulation of liver triglycerides in wild-type and AMPK α 2^{-/-} mice under the clamp conditions could be explained by hepatic lipid metabolism. In cultured hepatocytes isolated from mice following the different dietary treatments, activities of both fatty acid oxidation and de novo fatty acid synthesis were evaluated. The stimulatory effects of 5-aminoimidazole-4-carboxamide-1- β -D-ribofuranoside (AICAR), an AMPK activator, and insulin on fatty acid oxidation and synthesis are shown in Fig. 4A and B, respectively (for corresponding basal metabolic activities, see supplementary Table 3, available in an online appendix). In hepatocytes from both Chow and cHF diet-fed mice, the absence of AMPK α 2 was associated with a trend for lower AICAR-stimulated fatty acid oxidation. Although hepatocytes

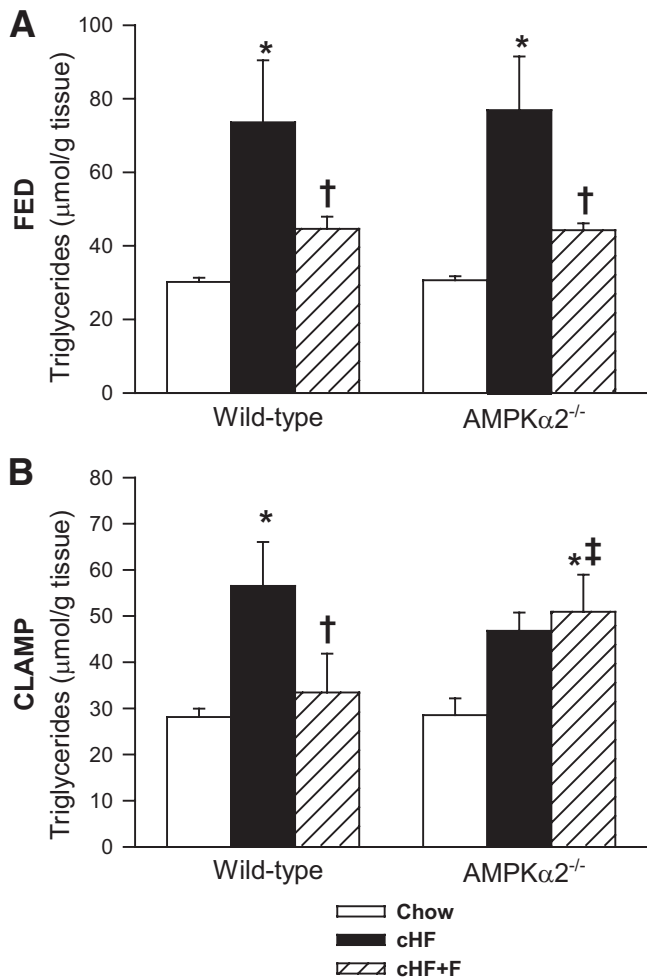


FIG. 3. Triglyceride concentration in the livers of ad libitum-fed mice (A) and mice killed at the end of a 3-h infusion period of the hyperinsulinemic-euglycemic clamp (B). Wild-type and AMPK α 2^{-/-} mice were fed either a Chow diet, cHF, or cHF+F for 9 weeks. The data are the means \pm SE (A, $n = 13$ –15; B, $n = 8$ –14). * $P < 0.05$ versus genotype Chow; † $P < 0.05$ versus genotype cHF; ‡ $P < 0.05$ versus wild-type on respective diet. For the detailed fatty acid composition of triglyceride fractions in the livers of ad libitum-fed mice, see supplementary Table 4.

from cHF-fed mice showed reduced stimulatory effect of AICAR irrespective of the genotype, cHF+F feeding normalized this defect in wild-type but not in AMPK α 2^{-/-} hepatocytes (Fig. 5A), suggesting AMPK-dependent induction of capacity for fatty acid oxidation by n-3 LC-PUFAs in the liver. The stimulatory effect of insulin on de novo fatty acid synthesis was reduced in hepatocytes from cHF-fed wild-type mice, whereas it was retained in the hepatocytes from cHF-fed AMPK α 2^{-/-} mice (Fig. 4B). cHF+F feeding tended to restore the stimulatory effect of insulin only in wild-type hepatocytes (Fig. 4B).

To further characterize hepatic effects of differential dietary treatment, the expression of selected genes was quantified in total RNA isolated from the livers of mice subjected to hyperinsulinemic-euglycemic clamp (Fig. 4C and D). Feeding cHF diet suppressed expression of lipogenic genes stearoyl-CoA desaturase (SCD-1) and SREBP-1c in all groups (except for SREBP-1c in AMPK α 2^{-/-} mice). This suppression was partially counteracted by cHF+F diet in wild-type but not AMPK α 2^{-/-} mice (Fig. 4C and D). Together with the de novo fatty acid synthesis data, these results further support the AMPK α 2-

dependent improvement of liver insulin sensitivity by n-3 LC-PUFAs.

Changes in hepatic diacylglycerol levels are associated with insulin-sensitizing effects of n-3 LC-PUFAs.

To identify factors predisposing animals to insulin resistance in an AMPK α 2-dependent manner, a detailed analysis of hepatic lipids in ad libitum-fed mice was performed. No major genotype-dependent differences in the contents of either ceramides or phospholipids were observed (supplementary Table 4). In contrast, hepatic content of diacylglycerols was affected in a genotype- and diet-dependent manner (Fig. 5A). Wild-type mice fed the cHF+F diet had lower diacylglycerol content than genotype-matched cHF diet-fed mice, while this effect of the cHF+F diet was not observed in AMPK α 2^{-/-} mice. Moreover, the analysis of fatty acid composition of the diacylglycerol fraction in the liver revealed that wild-type as well as AMPK α 2^{-/-} mice fed cHF diet were characterized by marked increase in the level of PUFA but not monounsaturated or saturated fatty acids (Fig. 5B, C, and D and supplementary Table 4). The increase in the PUFA content tended to be smaller in the wild-type compared with AMPK α 2^{-/-} mice (~ 1.7 -fold and ~ 2.2 -fold, respectively). Administration of n-3 LC-PUFAs completely prevented accumulation of hepatic polyunsaturated diacylglycerols in wild-type mice, whereas their level in the AMPK α 2^{-/-} animals, although decreased, was still significantly higher compared with genotype-matched Chow-fed mice (Fig. 5B). Regarding polyunsaturated diacylglycerols, α -linolenic acid (18:3n-3) appeared to be by far the most differentially regulated PUFA in the diacylglycerol fraction in the two genotypes (supplementary Table 4). In addition, cHF+F diet markedly reduced hepatic content of monounsaturated diacylglycerols in wild-type but not in knockout animals (Fig. 5C). Hepatic diacylglycerol levels and their fatty acid composition were also analyzed in mice subjected to hyperinsulinemic-euglycemic clamp (supplementary Table 5 and supplementary Fig. 3). No significant differences among the groups were observed in total diacylglycerols content or in their saturated or monounsaturated fatty acid fractions (supplementary Fig. 3).

DISCUSSION

Previous animal studies demonstrated that n-3 LC-PUFAs could counteract the development of both hepatic steatosis (8,18,36,37) and hepatic insulin resistance (8,9,16), while suppressing lipogenesis and augmenting lipid catabolism in the liver (8,13,19,21). Using mice with a whole-body deletion of AMPK α 2 and high-fat feeding, we show for the first time that AMPK α 2 is required for the effect of n-3 LC-PUFAs to preserve whole-body, muscle, and especially hepatic insulin sensitivity, as well as to suppress hepatic and plasma triglycerides as well as NEFA levels under hyperinsulinemic-euglycemic clamp conditions. In contrast, AMPK α 2 was not required for protection by n-3 LC-PUFAs from hepatic lipid accumulation and dyslipidemia in ad libitum-fed mice.

In addition to AMPK α 2, PPAR α was previously identified as an important determinant of n-3 LC-PUFA's effect on lipid metabolism, especially short-term modulation of hepatic gene expression (14) and insulin sensitivity (16). Thus, the reduction in hepatic triglyceride concentrations by fish oil feeding did not rescue insulin action in PPAR α -null mice, while hepatic diacylglycerol concentrations were decreased by fish oil in a PPAR α -dependent manner

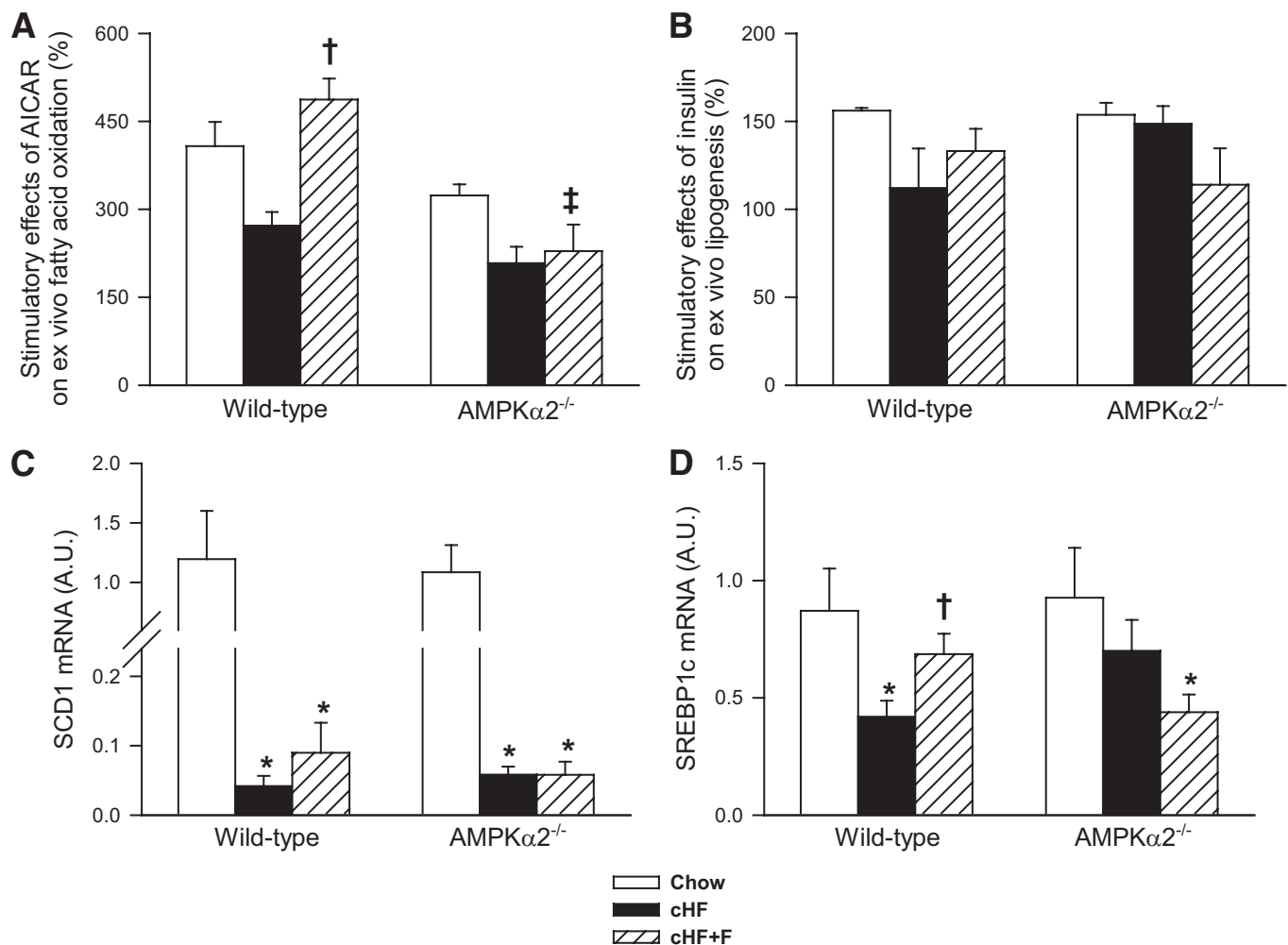


FIG. 4. The effect of differential dietary treatment on the regulation of metabolic fluxes in the liver. AICAR-stimulated fatty acid oxidation (A) and insulin-stimulated de novo fatty acid synthesis (B) in cultured hepatocytes isolated from wild-type and AMPK α 2^{-/-} mice fed for 9 weeks either a Chow diet, cHF, or cHF+F. For basal nonstimulated rates of lipid metabolism, see supplementary Table 3. The expression of SCD-1 (C) and SREBP-1c (D) genes was quantified in total RNA isolated from the livers of mice subjected to hyperinsulinemic-euglycemic clamp following the differential dietary treatment for 9 weeks. The data are means \pm SE (isolated hepatocytes, $n = 3$ in triplets; hepatic gene expression, $n = 5-8$). * $P < 0.05$ versus genotype Chow; † $P < 0.05$ versus genotype cHF; ‡ $P < 0.05$ versus wild-type on respective diet. A.U., arbitrary units.

and were associated with a preserved hepatic insulin sensitivity (16). It is generally accepted that 1) diacylglycerols rather than triglycerides or ceramides mediate hepatic insulin resistance in mice fed a high-fat diet (19,38,39), 2) diacylglycerol-induced insulin resistance depends on activation of protein kinase C, and 3) that polyunsaturated diacylglycerols in particular are better protein kinase C activators than saturated diacylglycerol species [reviewed in refs (38,39)]. Also our results showed that cHF diet-induced insulin resistance was associated primarily with the accumulation of PUFA in hepatic diacylglycerols and that n-3 LC-PUFA completely prevented cHF diet-induced increase in PUFA diacylglycerols in wild-type mice, whereas in AMPK α 2^{-/-} animals, the content of these lipids was still significantly higher compared with the control. Moreover, it was only in ad libitum-fed mice but not in mice subjected to hyperinsulinemic-euglycemic clamps that the levels of hepatic diacylglycerols and their fatty acid compositions were associated with hepatic insulin sensitivity. It is possible that under clamp conditions AMPK α 2-dependent effects of n-3 LC-PUFAs on liver diacylglycerols were masked by metabolic changes occurring during a 3-h infusion of insulin and glucose.

The failure of n-3 LC-PUFAs to decrease hepatic lipids in AMPK α 2^{-/-} mice under clamp conditions could be due to primary alterations in metabolic fluxes in the liver, reflecting 1) increased de novo fatty acid synthesis, 2) decreased secretion of VLDL triglycerides, or 3) reduced fatty acid oxidation. De novo fatty acid synthesis was not the responsible factor, because hepatocytes of the n-3 LC-PUFA-fed AMPK α 2^{-/-} mice showed decreased insulin-stimulated de novo fatty acid synthesis and reduced expression of SREBP-1c and SCD-1, as compared with hepatocytes isolated from n-3 LC-PUFA-fed wild-type mice, reflecting probably low insulin sensitivity of the liver in AMPK α 2^{-/-} mice. Moreover, AICAR-stimulated fatty acid oxidation in hepatocytes from n-3 LC-PUFA-fed AMPK α 2^{-/-} mice was markedly reduced as compared with those from wild-type mice, suggesting decreased hepatic capacity for fatty acid oxidation in the absence of AMPK α 2, which could contribute to enhanced lipid accumulation. Therefore, these experiments supported a major role of hepatic AMPK α 2 in the regulation of both insulin sensitivity and lipid metabolism by n-3 LC-PUFAs. However, the differential modulation of lipid accumulation by n-3 LC-PUFAs in the livers of wild-type and AMPK α 2^{-/-}

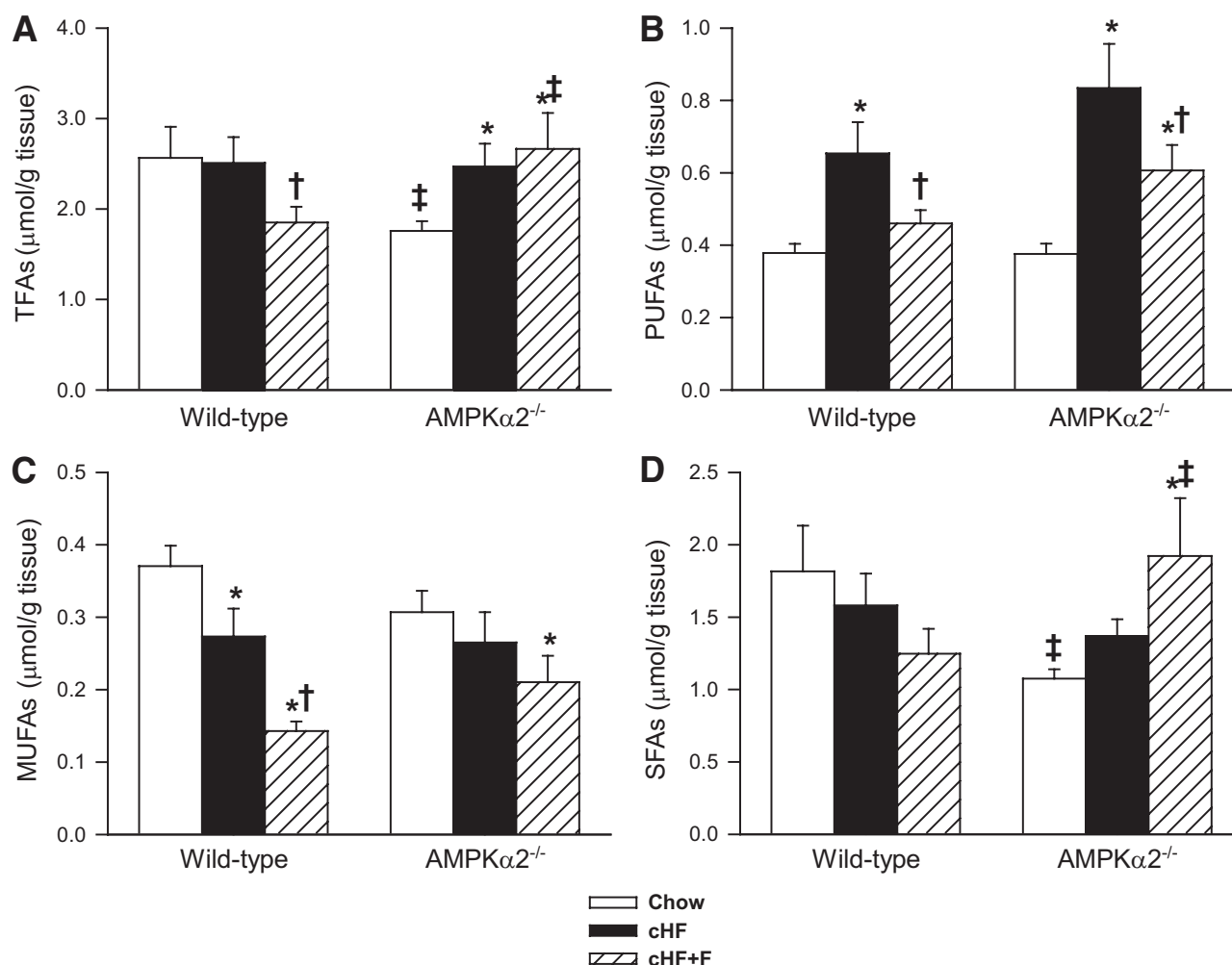


FIG. 5. The composition of fatty acids in hepatic diacylglycerol fraction in ad libitum-fed wild-type and $AMPK\alpha 2^{-/-}$ mice: total fatty acids (TFAs; *A*), PUFAs (*B*), monounsaturated fatty acids (MUFAs; *C*), and saturated fatty acids (SFAs; *D*). Animals were fed either a Chow diet, cHF, or cHF+F for 9 weeks. The data are the means \pm SE ($n = 13$ – 15). * $P < 0.05$ versus genotype Chow; † $P < 0.05$ versus genotype cHF; ‡ $P < 0.05$ versus wild-type on respective diet. For the detailed fatty acid composition of diacylglycerol fractions in the livers of ad libitum fed mice, see supplementary Table 4.

mice under clamp conditions could also be secondary to the $AMPK\alpha 2$ -dependent effects of n-3 LC-PUFAs in other tissues, resulting in a relatively high hepatic uptake of circulating NEFA in $AMPK\alpha 2^{-/-}$ mice. This is supported by persistently elevated plasma levels of NEFA in $AMPK\alpha 2^{-/-}$ mice, as well as by a significant correlation between plasma NEFA levels and hepatic triglyceride content observed under clamp conditions in $AMPK\alpha 2^{-/-}$ but not wild-type mice fed n-3 LC-PUFA-containing diet. Moreover, it has been shown in humans with nonalcoholic fatty liver disease that most of hepatic triglycerides arise from circulating NEFA (40). That plasma NEFA levels under clamp conditions were reduced only in wild-type but not in $AMPK\alpha 2^{-/-}$ mice fed n-3 LC-PUFAs may reflect a role of $AMPK\alpha 2$ in muscle lipid uptake mediated by lipoprotein lipase (41), as well as the antilipolytic effect of AMPK in adipose tissue, documented for $AMPK\alpha 1$ (42). In any case, decreased fatty acid oxidation in situ in the liver and, possibly even more importantly, abundant supply of circulating NEFA could be responsible for the lack of the antisteatotic effect of n-3 LC-PUFAs in $AMPK\alpha 2^{-/-}$ mice under clamp conditions (Fig. 6).

Previous studies reported contradictory results, showing either 1) activation of AMPK in rat liver (22) and

murine adipose tissue (24) or 2) no changes in AMPK activity in the liver, skeletal muscle, and heart of mice (43) in response to dietary n-3 LC-PUFAs. These discrepancies could be related to differences in dietary n-3 LC-PUFA intake, nutritional state of animals, and other parameters. In accordance with the involvement of $AMPK\alpha 2$ in various effects of n-3 LC-PUFAs, our results document activation of $AMPK\alpha 2$ (but not $AMPK\alpha 1$) in the liver of mice by long-term n-3 LC-PUFA treatment, in the absence of significant changes in either the AMP to ATP ratio assessed in whole liver extracts [not shown and ref (44)] or the phosphorylation status of LKB1, an upstream kinase for AMPK [not shown and ref (45)]. In addition, no effect on AMPK activity in either cultured hepatocytes (21) or embryonic kidney cells (not shown) of n-3 LC-PUFAs added to the cell culture medium could be detected. Therefore, the activation of $AMPK\alpha 2$ by n-3 LC-PUFAs probably does not depend on a direct interaction between n-3 LC-PUFAs and AMPK. On the other hand, induction of adiponectin by n-3 LC-PUFAs [results of this study and refs (46,47)] could be involved, because adiponectin activates AMPK in both the liver and skeletal muscle (35). Adiponectin is also required for the activation of AMPK upon administration of PPAR γ agonists thiazolidinedio-

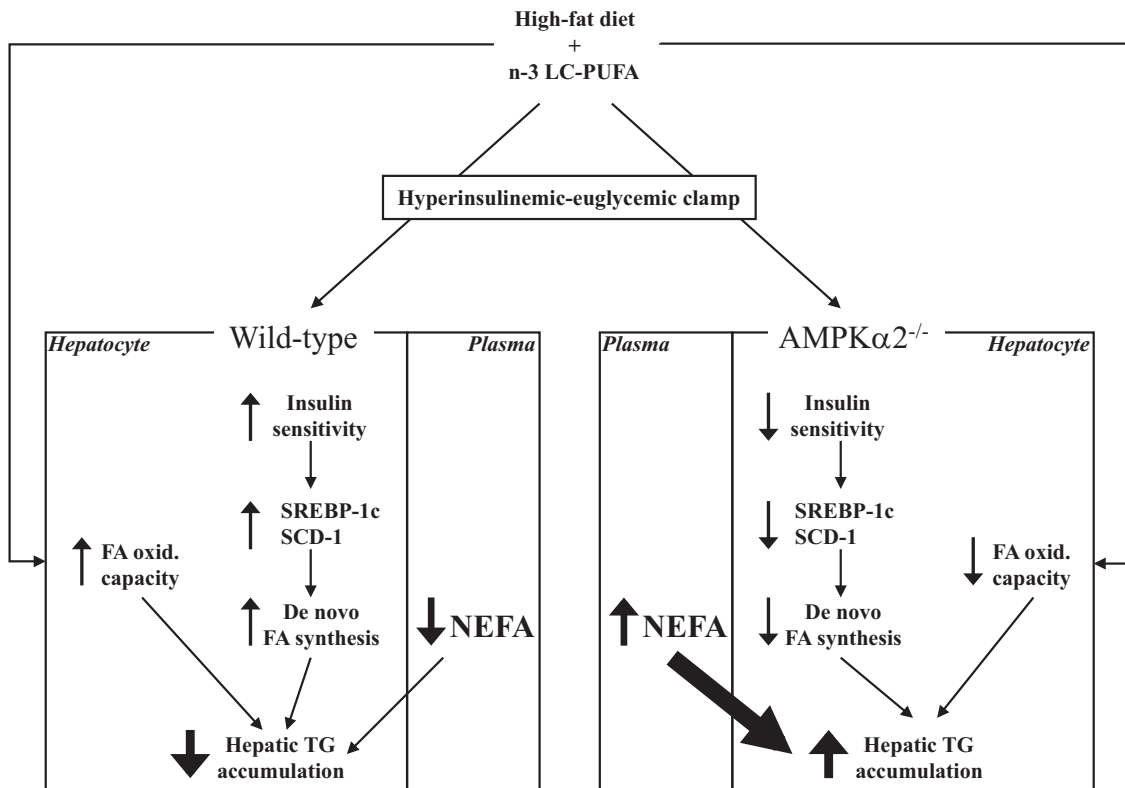


FIG. 6. Putative involvement of AMPK α 2 in antisteatotic action of n-3 LC-PUFAs in the liver. Dietary intake of n-3 LC-PUFAs increases the capacity of hepatocytes to oxidize fatty acids in wild-type (left panels) but not in AMPK α 2 $^{-/-}$ mice (right panels). When insulin and glucose levels are high, such as during hyperinsulinemic-euglycemic clamp, wild-type mice fed n-3 LC-PUFAs exhibit improved hepatic insulin sensitivity and decreased plasma levels of NEFAs as compared with high-fat diet-fed controls. This is associated with increased expression of lipogenic genes such as SREBP-1c and SCD-1 and increased drive for de novo fatty acid synthesis. Despite the elevated lipogenic drive under clamp conditions, the livers of wild-type mice fed n-3 LC-PUFAs show reduced accumulation of triglycerides. However, in AMPK α 2 $^{-/-}$ mice fed n-3 LC-PUFAs, hepatic triglyceride content is markedly elevated despite reduced rates of de novo fatty acid synthesis. This effect could be secondary to persisting elevated NEFA levels in circulation and thus better substrate availability in AMPK α 2 $^{-/-}$ mice under clamp conditions. FA, fatty acids; TG, triglycerides.

nes, whereas mice lacking adiponectin show decreased hepatic insulin sensitivity and reduced responsiveness to these compounds (48). Thus, absence of AMPK α 2 may blunt adiponectin-mediated effects of n-3 LC-PUFAs. In accordance with the previous study (49), plasma adiponectin levels tended to be reduced in AMPK α 2 $^{-/-}$ mice. Moreover, the induction of adiponectin by n-3 LC-PUFAs in AMPK α 2 $^{-/-}$ mice was compromised (Table 1 and supplementary Figure 2).

AMPK α 1 and AMPK α 2 contribute equally to total AMPK activity in the liver (50). In mice with liver-specific ablation of AMPK α 2 (27), hepatic AMPK α 2 was essential for suppressing hepatic glucose production and maintaining fasting blood glucose levels; however, the absence of AMPK α 2 did not affect inhibitory action of insulin on hepatic glucose production. In our study, although fasting blood glucose levels were unaltered by whole-body ablation of AMPK α 2, the beneficial effect of dietary n-3 LC-PUFAs on hepatic insulin sensitivity was clearly AMPK α 2-dependent. Differential regulation of glucose homeostasis in the above transgenic models likely reflects the complexity of whole-body (26) versus liver-specific (27) deletion of AMPK α 2. In contrast to the previous report, showing induction of adiposity and adipocyte hypertrophy in AMPK α 2 $^{-/-}$ mice fed a lard-based high-fat diet (49), our study documented a relatively low weight gain, low adiposity, and smaller fat cells in AMPK α 2 $^{-/-}$ mice fed a corn-oil based high-fat diet. This discrepancy could be related to the differences in the composition of experimental high-fat diets; however, our

results are consistent with the elevated sympathetic tone of AMPK α 2 $^{-/-}$ mice (26), which may stimulate energy dissipation in these animals. In any case, lower body weight of CHF-fed AMPK α 2 $^{-/-}$ mice as compared with their wild-type counterparts could be related to better insulin sensitivity of the former mice, as suggested by the differences in insulinemia, results of hyperinsulinemic-euglycemic clamp, stimulatory effect of insulin on lipogenesis, and expression of lipogenic genes in the liver.

In conclusion, the preservation of hepatic insulin sensitivity by n-3 LC-PUFAs in mice fed a high-fat diet depends on AMPK α 2. The accumulation of diacylglycerols, which is regulated in an AMPK α 2-dependent manner, could contribute to the modulation of hepatic insulin sensitivity in response to dietary n-3 LC-PUFAs. On the other hand, the AMPK α 2-dependent acute changes in lipid metabolism and hepatic triglyceride accumulation, which are unmasked under insulin-stimulated conditions such as during hyperinsulinemic-euglycemic clamp, largely reflect the extrahepatic action of n-3 LC-PUFAs. Our results are relevant for the development of novel strategies for prevention and treatment of the metabolic syndrome.

ACKNOWLEDGMENTS

This study was supported by grants from the Czech Science Foundation (301/10/1420) and the Ministry of Education, Youth and Sports (1M6837805002, COST BMB0602-OC08007) of the Czech Republic. Further sup-

port included grants from the European Commission (LSHM-CT-2004-005272, EXGENESIS) and from EPAX AS (Norway), as well as the research project AV0Z50110509. No other potential conflicts of interest relevant to this article were reported.

T.J. and M.R. performed experiments and wrote the manuscript. O.K., Z.M.J., D.M., V.K., M.H., and P.J. performed experiments. I.M. quantified tissue nucleotides. M.B. performed analysis of hepatic lipids and contributed to discussion. J.G. performed analysis of hepatic lipids. S.H. contributed to experiments on hepatocytes. T.E.J. contributed to analysis of muscle metabolism. P.F. introduced AMPK α 2^{-/-} mouse model to the Prague laboratory and contributed to discussion. S.H. performed experiments on kidney cells. B.V. developed AMPK α 2^{-/-} mice and contributed to discussion. J.K. wrote the manuscript.

The authors thank D. Grahame Hardie for the sheep AMPK α 1 and AMPK α 2 antibodies, Z. Szulc for the gift of *N*-palmitoyl-D-erythro-sphingosine (C17 base), and J. Jones (NMR Research Unit, Department of Biochemistry and Center for Neurosciences and Cell Biology, University of Coimbra, Coimbra, Portugal) for critical reading of the manuscript.

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Online-Only Appendix

TABLE 1. Blood glucose and plasma insulin levels during the insulin-stimulated (clamp) conditions and glucose turnover under basal (non-stimulated) conditions in wild-type and AMPK $\alpha 2^{-/-}$ mice

	Wild-type			AMPK $\alpha 2^{-/-}$		
	Chow	cHF	cHF+F	Chow	cHF	cHF+F
Blood glucose before clamp (mmol/l)	7.0 ± 0.8	8.0 ± 1.0	7.8 ± 0.8	8.5 ± 0.6	5.9 ± 0.1	7.4 ± 0.9
Steady-state blood glucose (mmol/l)	5.7 ± 0.5	5.7 ± 0.2	5.8 ± 0.2	5.8 ± 0.1	5.8 ± 0.7	5.9 ± 0.3
End-point plasma insulin (μ U/ml)	30 ± 10	29 ± 7	24 ± 8	21 ± 7	25 ± 7	28 ± 9
Basal glucose turnover (μ mol kg ⁻¹ min ⁻¹)	108 ± 11	86 ± 15	90 ± 15	118 ± 7	83 ± 23	104 ± 17

The data are presented as means ± SE (n = 5-8). Wild-type and AMPK $\alpha 2^{-/-}$ mice were fed either a Chow diet or corn oil-based high-fat diets without (cHF) or with 15% of the lipids in the form of n-3 LC-PUFA concentrate (cHF+F) for 9 weeks.

TABLE 2. Triglycerides, NEFA and cholesterol levels in plasma of wild-type and AMPK $\alpha 2^{-/-}$ mice under different metabolic conditions

	Wild-type			AMPK $\alpha 2^{-/-}$		
	Chow	cHF	cHF+F	Chow	cHF	cHF+F
Triglycerides (mmol/l)						
Fasted	0.46 ± 0.03	0.49 ± 0.03	0.46 ± 0.04	0.57 ± 0.05	0.44 ± 0.02	0.46 ± 0.02
Fed	1.17 ± 0.08	1.24 ± 0.11	0.62 ± 0.07*†	1.04 ± 0.06	1.22 ± 0.08	0.73 ± 0.07*†
Clamp	0.44 ± 0.05	0.64 ± 0.09	0.41 ± 0.04†	0.38 ± 0.06	0.55 ± 0.05	0.53 ± 0.08
NEFA (mmol/l)						
Fasted	1.16 ± 0.07	0.84 ± 0.08*	0.78 ± 0.05*	1.34 ± 0.05‡	0.89 ± 0.06*	0.87 ± 0.05*
Fed	0.90 ± 0.05	0.94 ± 0.05	0.59 ± 0.05*†	0.88 ± 0.04	0.99 ± 0.06	0.68 ± 0.04*†
Clamp	0.25 ± 0.02	0.39 ± 0.05	0.27 ± 0.03†	0.21 ± 0.03	0.41 ± 0.11	0.37 ± 0.07
Cholesterol (mmol/l)						
Fasted	1.77 ± 0.07	2.81 ± 0.22	2.00 ± 0.15†	1.78 ± 0.04	2.67 ± 0.12	2.02 ± 0.08†
Fed	2.25 ± 0.08	4.12 ± 0.25*	3.10 ± 0.20*†	2.10 ± 0.06	3.94 ± 0.18*	2.75 ± 0.14*†
Clamp	2.26 ± 0.13	2.98 ± 0.22	2.51 ± 0.15	1.83 ± 0.09	2.99 ± 0.24	2.63 ± 0.22

The data are presented as means ± SE (for fasted and fed mice, n = 13-15; for clamp, n = 8-14). Wild-type and AMPK $\alpha 2^{-/-}$ mice were fed either a Chow diet or corn oil-based high-fat diets without (cHF) or with 15% of the lipids in the form of n-3 LC-PUFA concentrate (cHF+F) for 9 weeks. Plasma parameters were assessed at 8 weeks (fasted values) or 9 weeks (fed and clamp values), respectively. * $P < 0.05$ vs. genotype Chow; † $P < 0.05$ vs. genotype cHF; ‡ $P < 0.05$ vs. wild-type on respective diet.

TABLE 3. The basal rates of lipid metabolism in isolated hepatocytes of wild-type and AMPK α 2^{-/-} mice

	Wild-type			AMPK α 2 ^{-/-}		
	Chow	cHF	cHF+F	Chow	cHF	cHF+F
Fatty acid oxidation (pmol/h/mg protein)	13 ± 2	8 ± 1*	9 ± 1*	15 ± 2	12 ± 1‡	6 ± 1*†
Lipogenesis (pmol/h/mg protein)	108 ± 9	74 ± 24	43 ± 10*†	79 ± 10	51 ± 6	34 ± 6*

The data are presented as means ± SE (n = 3 analyzed in triplets). Wild-type and AMPK α 2^{-/-} mice were fed either a Chow diet or corn oil-based high-fat diets without (cHF) or with 15% of the lipids in the form of n-3 LC-PUFA concentrate (cHF+F) for 9 weeks. **P* < 0.05 vs. genotype Chow; †*P* < 0.05 vs. genotype cHF; ‡*P* < 0.05 vs. wild-type on respective diet.

TABLE 4. The content and composition of hepatic lipids in wild-type and AMPK α 2^{-/-} mice killed in ad libitum fed state

	Wild-type			AMPK α 2 ^{-/-}		
	Chow	cHF	cHF+F	Chow	cHF	cHF+F
Total ceramide (nmol/g)	121.45 ± 9.33	119.71 ± 6.43	125.89 ± 14.66	123.34 ± 4.84	117.91 ± 7.53	123.82 ± 6.42
Triglyceride species						
Total fatty acids (μmol/g)	38.10 ± 3.18	116.55 ± 14.51*	80.35 ± 8.80†	50.31 ± 4.60	128.04 ± 18.59*	76.80 ± 6.63†
Myristic (14:0) (μmol/g)	0.38 ± 0.05	1.25 ± 0.22*	0.63 ± 0.08†	0.44 ± 0.04	1.27 ± 0.25*	0.59 ± 0.06†
Palmitic (16:0) (μmol/g)	11.12 ± 0.94	30.39 ± 4.10*	20.27 ± 2.88†	14.44 ± 1.06	35.25 ± 6.10*	17.17 ± 1.86†
Palmitoleic (16:1n7) (μmol/g)	2.82 ± 0.35	1.54 ± 0.46*	0.54 ± 0.10*	3.01 ± 0.37	1.35 ± 0.41*	0.81 ± 0.41*
Stearic (18:0) (μmol/g)	0.85 ± 0.09	1.90 ± 0.15*	1.60 ± 0.08*	0.98 ± 0.08	2.52 ± 0.31*‡	1.83 ± 0.14*†
Oleic (18:1n9) (μmol/g)	13.23 ± 1.22	27.05 ± 3.33	15.08 ± 1.72*†	16.68 ± 1.18	30.81 ± 5.84	15.52 ± 1.60*†
Linoleic (18:2n6) (μmol/g)	7.68 ± 0.61	47.52 ± 5.98*	25.98 ± 2.67*†	12.16 ± 2.60	49.22 ± 5.78*	25.81 ± 2.43*†
Arachidic (20:0) (μmol/g)	0.08 ± 0.01	0.23 ± 0.04*	0.18 ± 0.01*	0.12 ± 0.02	0.28 ± 0.05*	0.19 ± 0.02†
α-Linolenic (C18:3n3) (μmol/g)	0.43 ± 0.03	1.42 ± 0.24*	0.81 ± 0.09†	0.56 ± 0.06	1.52 ± 0.28*	0.79 ± 0.07†
Behenic (22:0) (nmol/g)	71.89 ± 5.67	89.37 ± 11.63	31.13 ± 2.65*†	81.32 ± 3.67	98.81 ± 15.97	43.53 ± 7.20*†
Arachidonic (20:4n6) (μmol/g)	0.56 ± 0.04	3.15 ± 0.58*	0.41 ± 0.04†	0.84 ± 0.12	3.72 ± 0.79*	0.47 ± 0.06†
Lignoceric (24:0) (nmol/g)	7.30 ± 0.80	10.28 ± 1.07	9.93 ± 1.04	8.14 ± 0.70	14.66 ± 3.10*	11.19 ± 0.89
Eicosapentaenoic (20:5n3) (μmol/g)	0.13 ± 0.01	0.25 ± 0.04	1.91 ± 0.21*†	0.13 ± 0.01	0.23 ± 0.05	1.33 ± 0.13*†‡
Nervonic (24:1n9) (nmol/g)	7.43 ± 1.01	8.23 ± 0.58	10.01 ± 0.89	6.94 ± 0.28	10.04 ± 1.30*	12.30 ± 1.17*
Docosahexaenoic (22:6n3) (μmol/g)	0.73 ± 0.06	1.74 ± 0.29	12.90 ± 1.08*†	0.84 ± 0.05	1.75 ± 0.31	12.21 ± 1.21*†
Diacylglycerol species						
Total fatty acids (μmol/g)	2.57 ± 0.34	2.51 ± 0.29	1.85 ± 0.18†	1.76 ± 0.11‡	2.47 ± 0.25*	2.66 ± 0.40*‡
Myristic (14:0) (μmol/g)	0.08 ± 0.01	0.08 ± 0.01	0.06 ± 0.01	0.06 ± 0.00	0.06 ± 0.01	0.06 ± 0.01

Palmitic (16:0) ($\mu\text{mol/g}$)	1.02 \pm 0.13	0.95 \pm 0.13	0.70 \pm 0.06	0.69 \pm 0.05	0.90 \pm 0.09	1.12 \pm 0.22‡
Palmitoleic (16:1n7) ($\mu\text{mol/g}$)	0.09 \pm 0.01	0.03 \pm 0.00*	0.01 \pm 0.00*	0.06 \pm 0.01‡	0.02 \pm 0.00*	0.02 \pm 0.01*
Stearic (18:0) ($\mu\text{mol/g}$)	0.70 \pm 0.18	0.54 \pm 0.10	0.48 \pm 0.11	0.31 \pm 0.02	0.40 \pm 0.04	0.65 \pm 0.16
Oleic (18:1n9) ($\mu\text{mol/g}$)	0.28 \pm 0.02	0.25 \pm 0.03	0.13 \pm 0.01*†	0.24 \pm 0.02	0.25 \pm 0.04	0.19 \pm 0.03
Linoleic (18:2n6) ($\mu\text{mol/g}$)	0.23 \pm 0.02	0.49 \pm 0.07*	0.27 \pm 0.03†	0.24 \pm 0.02	0.57 \pm 0.07*	0.36 \pm 0.05†
Arachidic (20:0) (nmol/g)	15.23 \pm 4.54	11.84 \pm 1.95	11.87 \pm 2.85	7.67 \pm 0.56	9.47 \pm 0.62	10.94 \pm 0.92
α -Linolenic (18:3n3) (nmol/g)	12.61 \pm 1.52	12.60 \pm 1.88	6.55 \pm 0.50*†	7.64 \pm 0.64‡	10.73 \pm 1.52	13.23 \pm 2.62‡
Arachidonic (20:4n6) (nmol/g)	72.11 \pm 3.89	83.54 \pm 5.32	30.86 \pm 4.35*†	65.43 \pm 4.89	100.95 \pm 16.46*	37.75 \pm 4.28*†
Eicosapentaenoic (20:5n3) (nmol/g)	10.43 \pm 0.75	8.59 \pm 0.89	19.97 \pm 1.62*†	8.58 \pm 0.58	8.37 \pm 8.84	20.29 \pm 1.27*†
Nervonic (24:1n9) (nmol/g)	0.80 \pm 0.10	1.36 \pm 0.32	1.18 \pm 0.12	0.83 \pm 0.06	1.38 \pm 0.37	1.38 \pm 0.21
Docosahexaenoic (22:6n3) ($\mu\text{mol/g}$)	0.05 \pm 0.00	0.06 \pm 0.01	0.13 \pm 0.01*†	0.06 \pm 0.01	0.14 \pm 0.05*‡	0.17 \pm 0.02*
Phospholipid species						
Total fatty acids ($\mu\text{mol/g}$)	67.96 \pm 1.61	65.83 \pm 1.09	70.75 \pm 1.39†	68.20 \pm 0.97	67.62 \pm 1.40	74.45 \pm 2.44*†
Myristic (14:0) ($\mu\text{mol/g}$)	0.14 \pm 0.01	0.22 \pm 0.08	0.16 \pm 0.01	0.26 \pm 0.08	0.15 \pm 0.02	0.15 \pm 0.01
Palmitic (16:0) ($\mu\text{mol/g}$)	18.38 \pm 0.46	13.80 \pm 0.24*	17.38 \pm 0.21†	18.16 \pm 0.30	14.18 \pm 0.34*	18.20 \pm 0.87†
Palmitoleic (16:1n7) ($\mu\text{mol/g}$)	1.40 \pm 0.12	0.17 \pm 0.01*	0.25 \pm 0.11*	1.36 \pm 0.06	0.17 \pm 0.01*	0.33 \pm 0.19*
Stearic (18:0) ($\mu\text{mol/g}$)	11.21 \pm 0.36	14.39 \pm 0.28*	13.53 \pm 0.47*	11.56 \pm 0.25	14.81 \pm 0.33*	14.30 \pm 0.36*
Oleic (18:1n9) ($\mu\text{mol/g}$)	5.23 \pm 0.21	3.34 \pm 0.14*	3.80 \pm 0.21*	5.02 \pm 0.14	3.42 \pm 0.14*	4.11 \pm 0.36*†
Linoleic (18:2n6) ($\mu\text{mol/g}$)	12.37 \pm 0.29	14.27 \pm 0.39*	14.89 \pm 0.42*	12.00 \pm 0.21	14.38 \pm 0.45*	15.91 \pm 0.41*†
Arachidic (20:0) ($\mu\text{mol/g}$)	0.10 \pm 0.01	0.11 \pm 0.01	0.15 \pm 0.01*†	0.10 \pm 0.00	0.11 \pm 0.01	0.15 \pm 0.01*†
α -Linolenic (C18:3n3) ($\mu\text{mol/g}$)	0.07 \pm 0.01	0.04 \pm 0.00*	0.05 \pm 0.00	0.07 \pm 0.01	0.05 \pm 0.00*	0.06 \pm 0.01
Behenic (22:0) ($\mu\text{mol/g}$)	0.13 \pm 0.01	0.03 \pm 0.01*	0.04 \pm 0.01*	0.13 \pm 0.01	0.02 \pm 0.00*	0.04 \pm 0.02*
Arachidonic (20:4n6) ($\mu\text{mol/g}$)	11.83 \pm 0.66	13.70 \pm 0.30*	5.60 \pm 0.47*†	12.85 \pm 0.18	13.75 \pm 0.79*	6.62 \pm 0.80†
Lignoceric (24:0) (nmol/g)	9.90 \pm 1.00	15.69 \pm 6.14	11.32 \pm 0.70	13.82 \pm 3.82	10.46 \pm 0.72	10.29 \pm 0.83
Eicosapentaenoic (20:5n3) ($\mu\text{mol/g}$)	0.44 \pm 0.08	0.08 \pm 0.01*	2.01 \pm 0.17*†	0.27 \pm 0.01	0.16 \pm 0.10	1.78 \pm 0.19*†
Docosahexaenoic (22:6n3) ($\mu\text{mol/g}$)	6.64 \pm 0.39	5.64 \pm 0.18	12.88 \pm 0.65*†	6.42 \pm 0.13	6.40 \pm 0.53	12.80 \pm 0.75*†

The data are the means \pm SE (n = 8-14). Wild-type and AMPK α 2^{-/-} mice were fed either a Chow diet or corn oil-based high-fat diets without (cHF) or with 15% of the lipids in the form of n-3 LC-PUFA concentrate (cHF+F) for 9 weeks and killed in ad libitum fed state. **P* < 0.05 vs. genotype Chow; †*P* < 0.05 vs. genotype cHF; ‡*P* < 0.05 vs. wild-type on respective diet.

The intake of n-3 LC-PUFA resulted in a marked increase in EPA and DHA concentration in all lipid fractions, independently of the genotype and with relatively small changes in the diacylglycerol fraction. Except for EPA and DHA (and a relatively low content of arachidonic acid and high content of nervonic acid), levels of individual fatty acids in the triglyceride fraction reflected the total tissue content of triglycerides. The fatty acid profile of the phospholipid fraction was only marginally affected by either diet or genotype. In diacylglycerol fraction, the content of several fatty acids (i.e., palmitic, palmitoleic, oleic, and α -linolenic acid) was differentially affected by the cHF+F diet, depending on the genotype and explaining the effect of the cHF+F diet on total hepatic content of diacylglycerols (see Fig. 5A of the main text). α -linolenic acid was by far the most differentially affected fatty acid, with a ~1.9-fold decrease in response to cHF+F diet in wild-type mice, and a ~1.2-increase produced by the same diet in AMPK α 2^{-/-} mice. This fatty acid also contributed to the lower content of liver diacylglycerols in AMPK α 2^{-/-} compared to wild-type mice fed the Chow diet.

TABLE 5. The content and composition of hepatic diacylglycerols in wild-type and AMPK α 2^{-/-} mice subjected to hyperinsulinemic-euglycemic clamp

	Wild-type			AMPK α 2 ^{-/-}		
	Chow	cHF	cHF+F	Chow	cHF	cHF+F
Diacylglycerol species						
Total fatty acids (μ mol/g)	1.90 \pm 0.21	2.43 \pm 0.32	2.57 \pm 0.32	1.99 \pm 0.12	2.72 \pm 0.35	2.43 \pm 0.48
Myristic (14:0) (μ mol/g)	0.06 \pm 0.01	0.05 \pm 0.01	0.07 \pm 0.01	0.05 \pm 0.00	0.06 \pm 0.01	0.06 \pm 0.01
Palmitic (16:0) (μ mol/g)	0.58 \pm 0.07	0.67 \pm 0.09	0.85 \pm 0.17	0.62 \pm 0.03	0.73 \pm 0.08	0.75 \pm 0.15
Palmitoleic (16:1n7) (μ mol/g)	0.03 \pm 0.00	0.01 \pm 0.00*	0.01 \pm 0.00*	0.03 \pm 0.00	0.01 \pm 0.00*	0.01 \pm 0.00*
Stearic (18:0) (μ mol/g)	0.53 \pm 0.18	0.41 \pm 0.05	0.68 \pm 0.19	0.42 \pm 0.05	0.48 \pm 0.10	0.68 \pm 0.26
Oleic (18:1n9) (μ mol/g)	0.24 \pm 0.04	0.29 \pm 0.05	0.22 \pm 0.04	0.31 \pm 0.05	0.33 \pm 0.05	0.21 \pm 0.04
Linoleic (18:2n6) (μ mol/g)	0.29 \pm 0.05	0.81 \pm 0.15*	0.49 \pm 0.09	0.35 \pm 0.04	0.91 \pm 0.14*	0.47 \pm 0.08*†
Arachidic (20:0) (nmol/g)	13.16 \pm 4.54	9.63 \pm 0.72	14.52 \pm 3.47	9.63 \pm 1.08	11.94 \pm 2.51	16.70 \pm 6.43
α -Linolenic (18:3n3) (nmol/g)	17.76 \pm 5.65	13.88 \pm 3.53	13.70 \pm 4.46	11.91 \pm 1.68	13.03 \pm 2.17	11.09 \pm 2.80
Arachidonic (20:4n6) (nmol/g)	83.84 \pm 14.03	68.67 \pm 2.07	33.82 \pm 3.17*†	106.37 \pm 11.29	80.26 \pm 4.40*	39.60 \pm 4.58*†
Lignoceric (24:0) (nmol/g)	1.29 \pm 0.21	1.09 \pm 0.12	1.56 \pm 0.23	1.21 \pm 0.08	1.02 \pm 0.13	1.08 \pm 0.21
Eicosapentaenoic (20:5n3) (nmol/g)	5.73 \pm 0.45	5.10 \pm 0.75	11.96 \pm 0.99*†	6.26 \pm 0.82	5.40 \pm 0.58	12.48 \pm 1.12*†
Nervonic (24:1n9) (nmol/g)	11.63 \pm 2.56	9.83 \pm 2.32	11.61 \pm 2.24	10.87 \pm 2.25	12.37 \pm 2.85	11.64 \pm 1.82
Docosahexaenoic (22:6n3) (μ mol/g)	0.05 \pm 0.01	0.07 \pm 0.01	0.16 \pm 0.03*†	0.07 \pm 0.01	0.08 \pm 0.01	0.16 \pm 0.04*

The data are the means \pm SE (n = 5-8). Wild-type and AMPK α 2^{-/-} mice were fed either a Chow diet or corn oil-based high-fat diets without (cHF) or with 15% of the lipids in the form of n-3 LC-PUFA concentrate (cHF+F) for 9 weeks. Liver tissue was collected immediately after the completion of hyperinsulinemic-euglycemic clamp. **P* < 0.05 vs. genotype Chow; †*P* < 0.05 vs. genotype cHF; ‡*P* < 0.05 vs. wild-type on respective diet.

RESEARCH DESIGN AND METHODS

Animals and treatments. Whole-body AMPK $\alpha 2^{-/-}$ mice and wild-type littermate controls generated on a hybrid C57BL/6 and 129/Sv genetic background (1) and backcrossed to C57BL/6J mice for nine generations were used. At four weeks of age, mice were weaned onto a standard laboratory Chow (lipid content ~3.4% wt/wt; extruded R/M-H diet; Ssniff Spezialdiäten, Soest, Germany) and maintained at 22°C on a 12 h light–dark cycle (light on from 6 a.m.) with free access to food and water.

Two separate experiments were performed, in which four-month-old wild-type and AMPK $\alpha 2^{-/-}$ mice (n = 13–15), caged in groups of 3–4 mice, were fed the following diets for nine weeks: (i) Chow; (ii) a corn oil-based high-fat diet (cHF; lipid content ~35.2% wt/wt); or (iii) a cHF diet supplemented with n-3 LC-PUFA concentrate (46% DHA, 14% EPA; product EPAX 1050 TG; EPAX, a.s., Lysaker, Norway) replacing 15% of dietary lipids (cHF+F). The macronutrient and fatty acid composition of all diets has been described before (2). Fresh rations of food were distributed every two days, while food consumption and body weights were recorded once a week. Mice from the first experiment in ad libitum fed state were first anesthetized by diethylether, exsanguinated through the cervical incision, then sacrificed by cervical dislocation (between 8 a.m. and 10 a.m.), and EDTA-plasma, liver, quadriceps muscles, epididymal and subcutaneous (dorsolumbar) adipose tissues were collected for various analyses (see below). When indicated, plasma samples were also collected after eight weeks from mice fasted for 15 hours (between 8 a.m. and 11 p.m.). In order to assess insulin sensitivity, mice from the second experiment were studied under hyperinsulinemic-euglycemic conditions.

Real-time RT-PCR analysis. Total RNA was isolated from liver samples (stored in RNAlater Solution; Ambion, Austin, TX) homogenized in TRIzol Reagent (Invitrogen, Carlsbad, CA). A single-strand cDNA was synthesized from 0.5 μ g of total RNA and gene expression was assessed by real-time PCR, using the LightCycler® 480 Instrument and the kit LightCycler® 480 SYBR Green I (Roche Diagnostics, Mannheim, Germany). Transcript levels were normalized according to the expression levels of eukaryotic elongation factor 1a (ELF-1a). The following oligonucleotide primers, designed by Lasergene software (DNASar, Madison, WI), were used:

SCD1:

sense, 5'-ACTGGGGCTGCTAATCTCTGGGTGTA,
antisense, 3'-GGCTTTATCTCTGGGGTGGGTTTGTTA;

SREBP-1c:

sense, 5'-TACCCGTCCGTGTCCCCCTTTTC,
antisense, 3'-TGCGCTTCTACCCACGGCTCTG;

ELF-1a:

sense 5'-TGACAGCAAAAACGACCCACCAAT,
antisense, 3'-GGCCATCTTCCAGCTTCTTACCA.

Activity of $\alpha 1$ and $\alpha 2$ AMPK isoforms. Livers were collected by freeze-clamping. AMPK was immunoprecipitated from tissue extracts and the activity was assayed using a peptide substrate (3). AMPK $\alpha 1$ or AMPK $\alpha 2$ isoforms were immunoprecipitated using specific antibodies (gift of Dr. D. Grahame Hardie), which were bound to Protein G Sepharose (Amersham Pharmacia Biotech, Little Chalfont, UK). Immunoprecipitates were mixed with AMP (1 mmol/l), [γ -

^{32}P]ATP (1 mmol/l) and AMARA peptide (1 mmol/l; Vidia, Prague, Czech Republic) dissolved in HEPES-Brij buffer. Reaction proceeded for 30 min at 30°C and it was stopped by dropping the filter spotted with a reaction mixture into 1% phosphoric acid solution. The activity of 1 U/mg protein corresponds to 1 nmol ^{32}P -AMARA peptide/mg protein per min.

The analysis of blood samples from hyperinsulinemic-euglycemic clamps. To assess D-[3- ^3H]glucose, $^3\text{H}_2\text{O}$ and total glucose concentrations, blood samples were firstly deproteinized by precipitation with a $\text{ZnSO}_4/\text{Ba}(\text{OH})_2$, followed by centrifugation (4). The first aliquot of the supernatant was evaporated to dryness to determine the radioactivity corresponding to D-[3- ^3H]glucose. The second aliquot was used to determine the radioactivity of both D-[3- ^3H]glucose and $^3\text{H}_2\text{O}$. Plasma $^3\text{H}_2\text{O}$ radioactivity was then calculated as the difference between radioactivity in the second and the first aliquot. In the third aliquot of the supernatant, the total glucose concentration was assessed by the glucose oxidase method (Glukosa God 1500, PLIVA-Lachema, Czech Republic). The D-[3- ^3H]glucose specific activity was calculated by dividing the D-[3- ^3H]glucose enrichment of plasma (dpm/ml) by the total plasma glucose concentration (mg/ml). Time points, in which the steady-state D-[3- ^3H]glucose specific activity varied more than $\pm 15\%$, were not taken into account (4). The rates of insulin-stimulated glucose turnover (GTO) were determined as the ratio of the [3- ^3H]glucose infusion rate (dpm/min) to the specific activity of plasma glucose (5). The rate of glycogen synthesis in quadriceps muscle was quantified by measuring the incorporation of D-[3- ^3H]glucose into glycogen and calculated by dividing the radioactivity of D-[3- ^3H]glucosyl units in glycogen (dpm/kg) by the mean specific activity of D-[3- ^3H]glucose in plasma during the last hour of the clamp.

The rate of lipogenesis in isolated hepatocytes. De novo lipogenesis was assessed using ^{14}C -Acetate (1uCi/ml) (PerkinElmer, USA). Cells were incubated in 2 ml of a medium (M199, Gibco) containing ^{14}C acetate with or without 25 mM glucose and 100 nM insulin (Actrapid MC, Novoindustri, Denmark). After 2 hr of incubation at 37 °C (5% CO_2 and 95% O_2) cells were washed by PBS and harvested into 600 ul KOH (shaking at 70°C). ^{14}C incorporation into saponifiable FA was estimated (6) and expressed as pmol of acetate converted to fatty acid/mg protein per hour. Identical incubations in parallel wells without radioactivity were performed to determine protein concentrations.

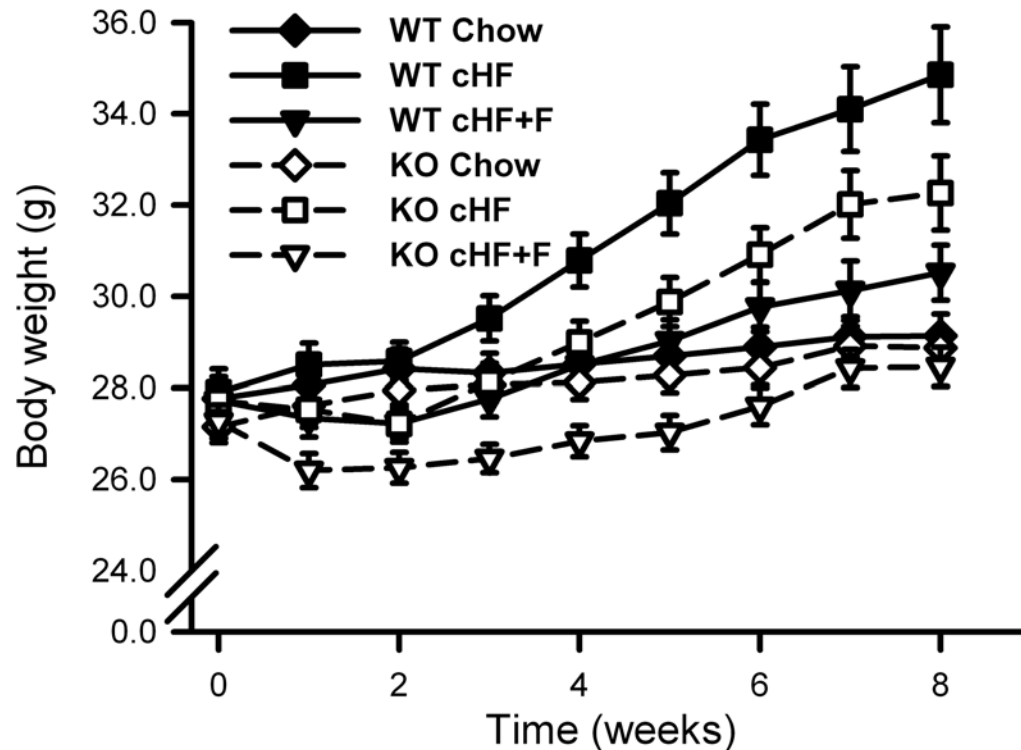
The rate of fatty acid oxidation in isolated hepatocytes. Palmitate oxidation was assessed using [1- ^{14}C] palmitate (PerkinElmer, USA). Cells were incubated for 45 min at 37°C in 800 ul of a medium (M199) supplemented with 1% BSA, 50 μM cold palmitate and [1- ^{14}C] palmitate (0,38 μCi /reaction) with or without 1 mM AICAR. The reactions were terminated by aspiration of the media, cells were washed by PBS and then incubated in 800 ul of 5 % perchloric acid for 15 min at room temperature. Palmitate oxidation was determined by measuring production of ^{14}C -labeled acid-soluble metabolites (ASM), a measure of tricarboxylic acid cycle intermediates and acetyl esters. The ASM were assessed in supernatants of the acid precipitate. Identical incubations in parallel wells without radioactivity were conducted to determine protein concentrations.

Total content and fatty acid composition of phospholipid, diacylglycerol, triglyceride, and ceramide fractions. Liver samples were pulverized in an aluminum mortar precooled in liquid nitrogen. The powder was transferred to a tube containing ice-cold methanol and 0.01%

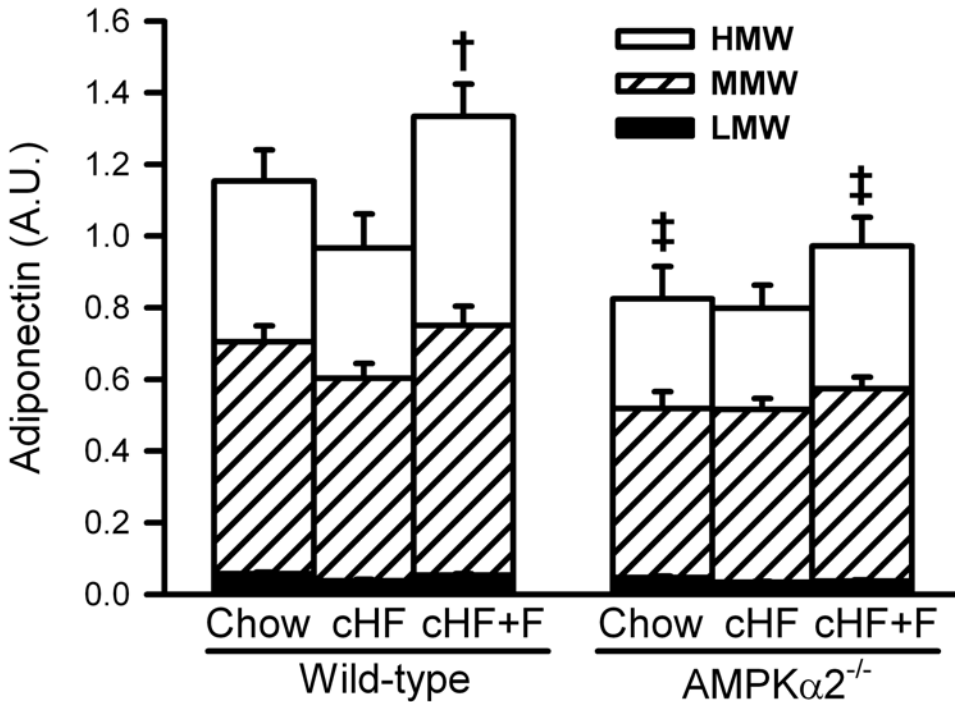
butylated hydroxytoluene (Sigma) as an antioxidant. Internal standards (1,2-diheptadecanoin, triheptadecanoin and 1,2-diheptadecanoyl-sn-glycero-3-phosphatidylcholine, Larodan Fine Chemicals, Sweden) were then added to the samples and lipids were extracted by the method of Folch. The fractions of total phospholipids, triglycerides and diacylglycerols were separated by thin-layer chromatography (7). Lipid class standards were spotted on the outside lanes of the chromatography plate to enable localization of the sample lipid classes. The gel bands corresponding to the standards were scraped off the plates, transferred to fresh tubes and then transmethylated in 14% methanolic boron trifluoride (Sigma) at 100°C for either 10 or 30 minutes (phospholipids and triglycerides). The content of resulting fatty acid methyl esters was determined by means of gas-liquid chromatography (8). The content of ceramides was determined as described previously (9) with the exception that N-palmitoyl-D-erythro-sphingosine (C17 Base; a gift from Dr. Zdzislaw Szulc, Medical University of South Carolina) was used as an internal standard.

FIGURE LEGENDS

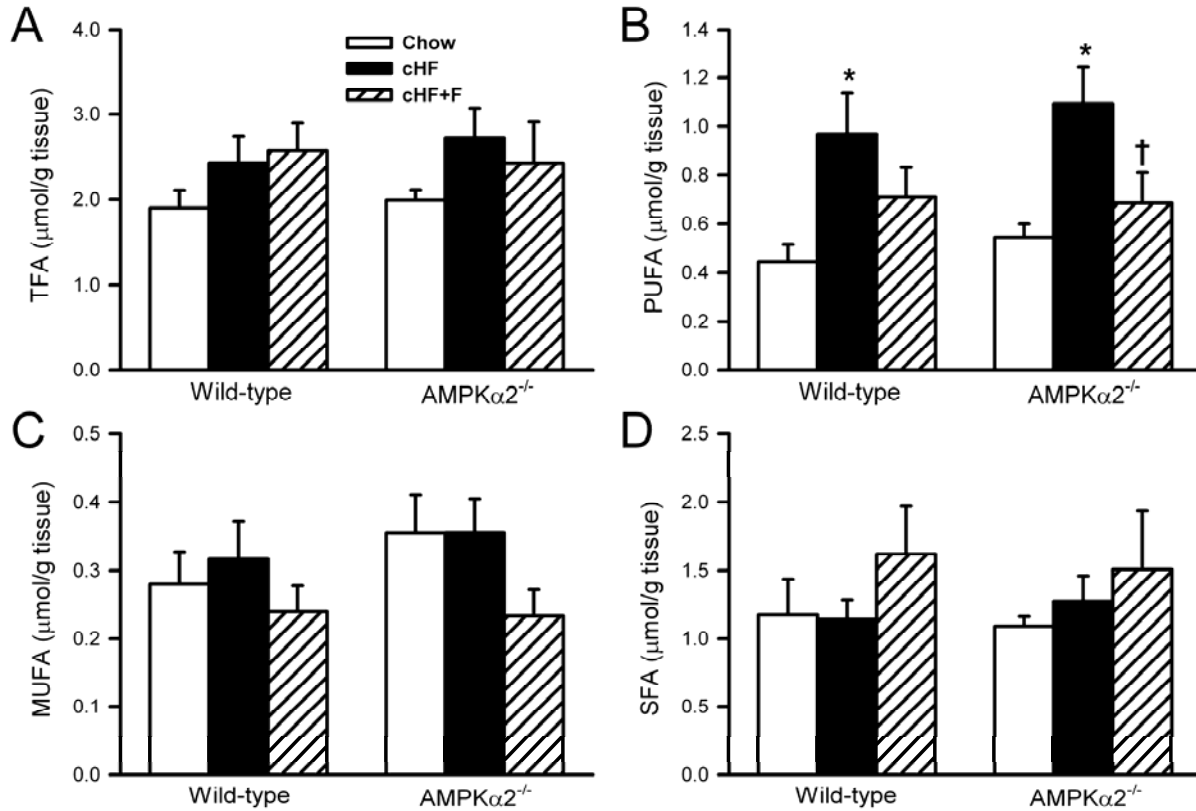
Supplemental Figure. 1. Body weight curves of wild-type (WT) and AMPK $\alpha 2^{-/-}$ (KO) mice fed either a Chow diet or corn oil-based high-fat diets without (cHF) or with 15% of the lipids in the form of n-3 LC-PUFA concentrate (cHF+F) for 9 weeks. The data are the means \pm SE (n = 27-30).



Supplemental Fig. 2. Adiponectin levels in plasma of wild-type and AMPK α 2^{-/-} mice fed either a Chow diet or corn oil-based high-fat diets without (cHF) or with 15% of the lipids in the form of n-3 LC-PUFA concentrate (cHF+F) for 9 weeks, and killed in ad libitum fed state. The total adiponectin levels and the distribution of adiponectin multimeric complexes were determined using Western blotting (10). The data are the means \pm SE (n = 13-15). **P* < 0.05 vs. genotype Chow; †*P* < 0.05 vs. genotype cHF; ‡*P* < 0.05 vs. wild-type on respective diet. Significance evaluated for the total adiponectin levels. A.U., arbitrary units; LMW – low molecular weight; MMW – medium molecular weight; HMW – high molecular weight.



Supplemental Fig. 3. The composition of fatty acids in hepatic diacylglycerol fraction in wild-type and AMPK $\alpha 2^{-/-}$ mice subjected to hyperinsulinemic-euglycemic clamp: total fatty acids (TFA; A), polyunsaturated fatty acids (PUFA; B), monounsaturated fatty acids (MUFA; C), saturated fatty acids (SFA; D). Animals were fed either a Chow diet or corn oil-based high-fat diets without (cHF) or with 15% of the lipids in the form of n-3 LC-PUFA concentrate (cHF+F) for 9 weeks, and then subjected to hyperinsulinemic-euglycemic clamp. Liver tissue was collected immediately after the completion of the clamp. Data are means \pm SE (n = 5-8). * P < 0.05 vs. genotype Chow; † P < 0.05 vs. genotype cHF.



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